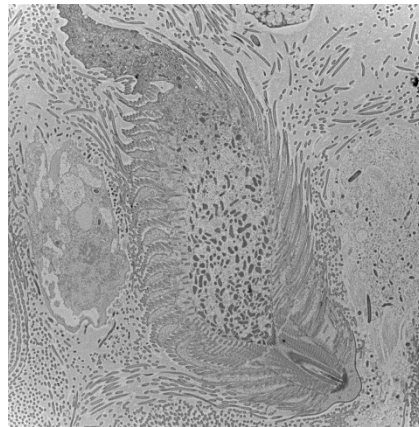


Assessment of subterranean termite symbiotic fauna under different diets

Sónia Duarte^{1,2}, Lina Nunes^{1,2}, Paulo A.V. Borges², Carl G. Fossdal³

COST-STSM-FP1303-20909



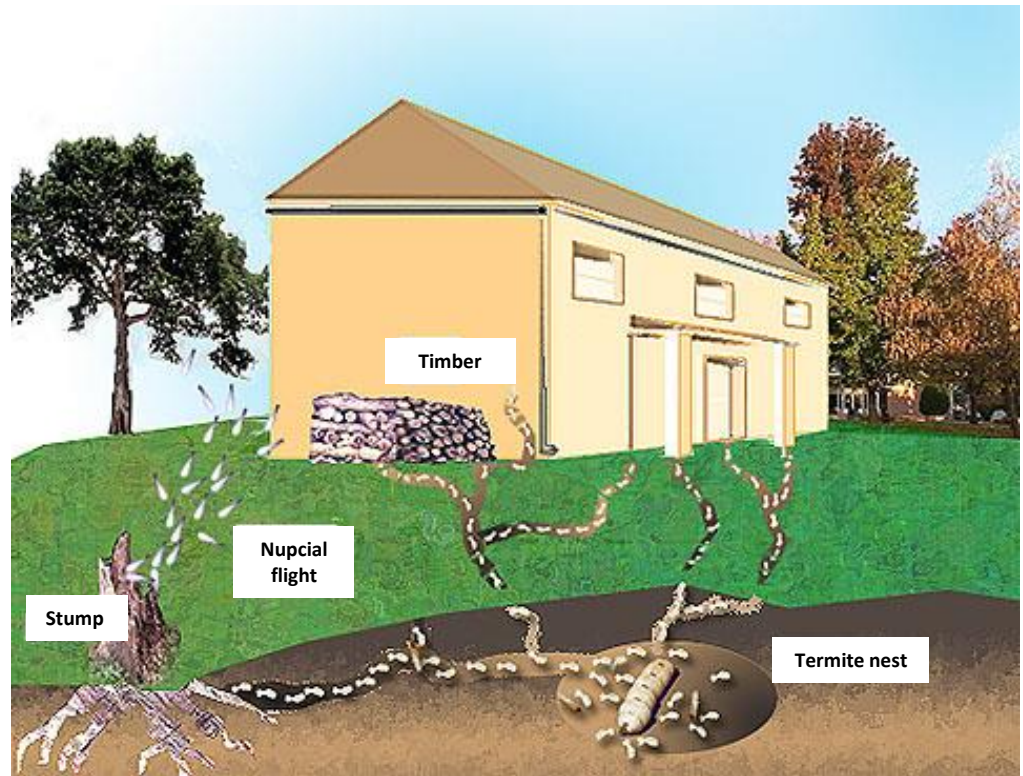
1

2

3

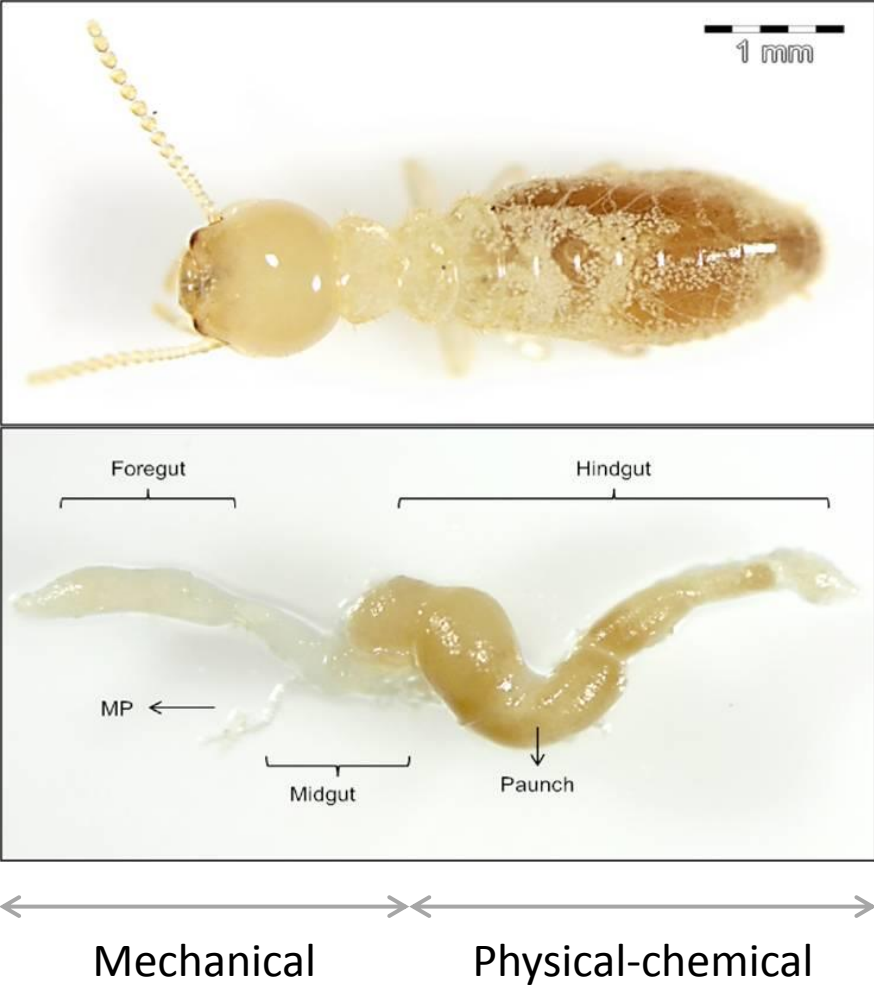
Why termites?

Attacking wood in service... and also biobased materials of cellulosic base

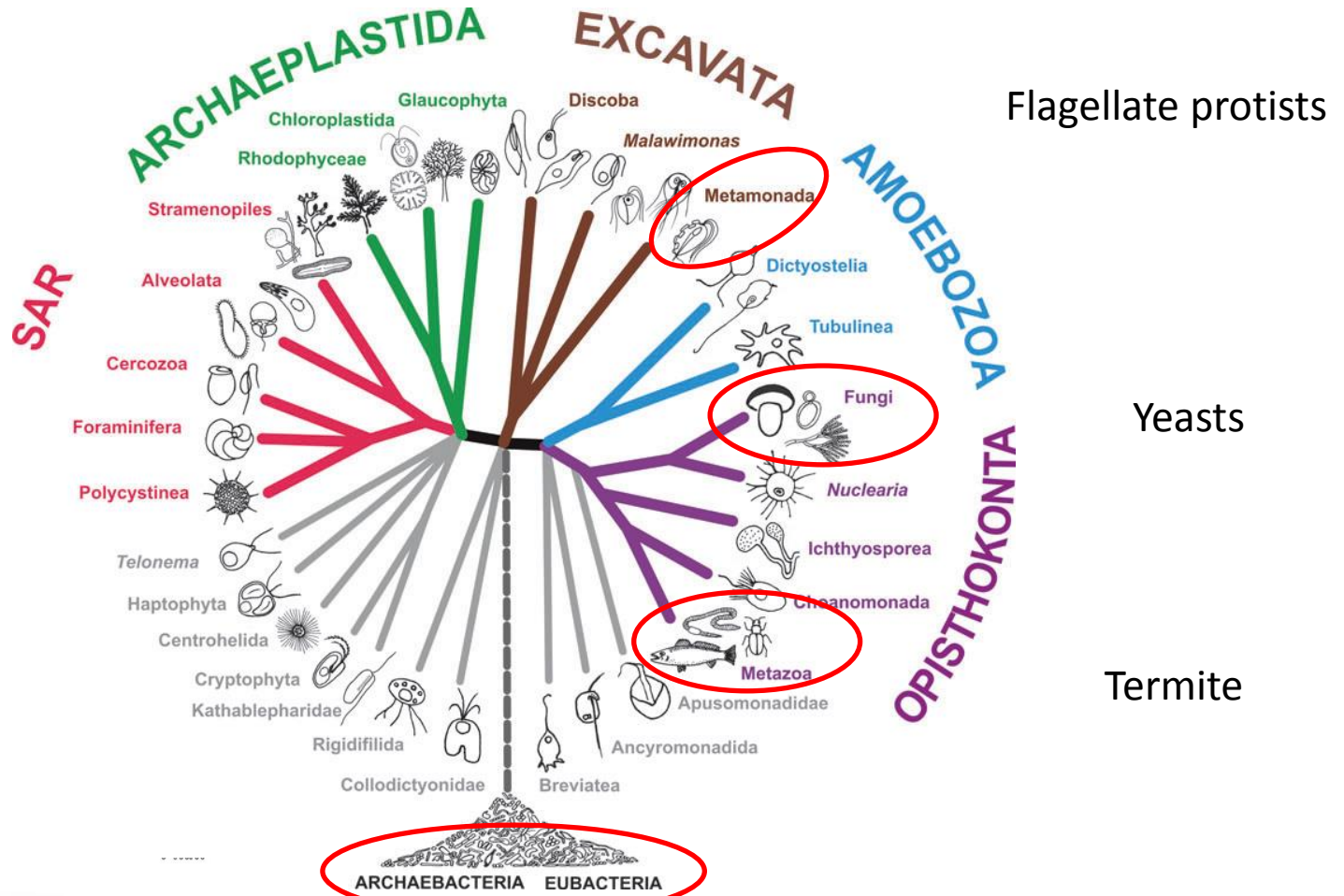


Important in natural ecosystems, worrying in human made environments

How do subterranean termites digest lignocellulose?



Who are part of the subterranean termite holobiont?



Flagellate protists

Yeasts

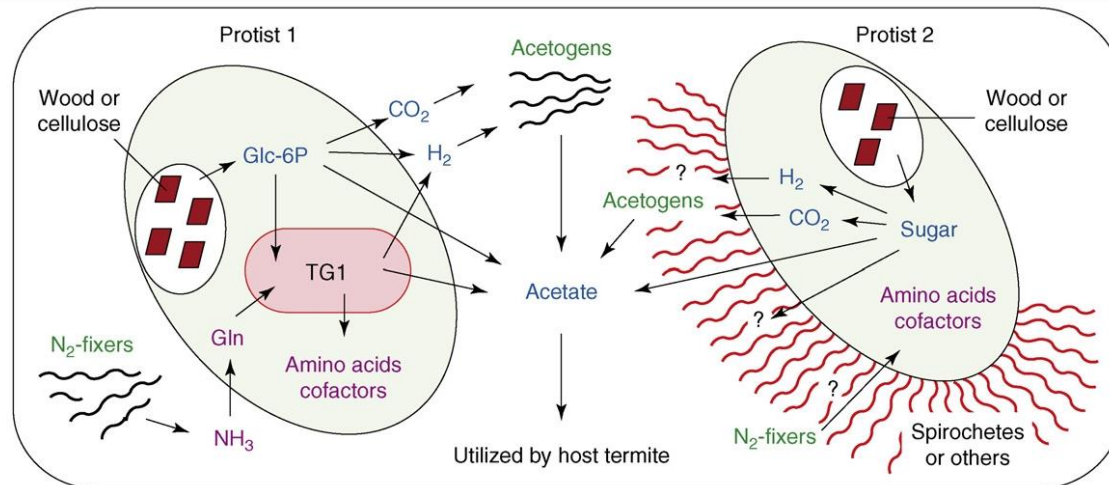
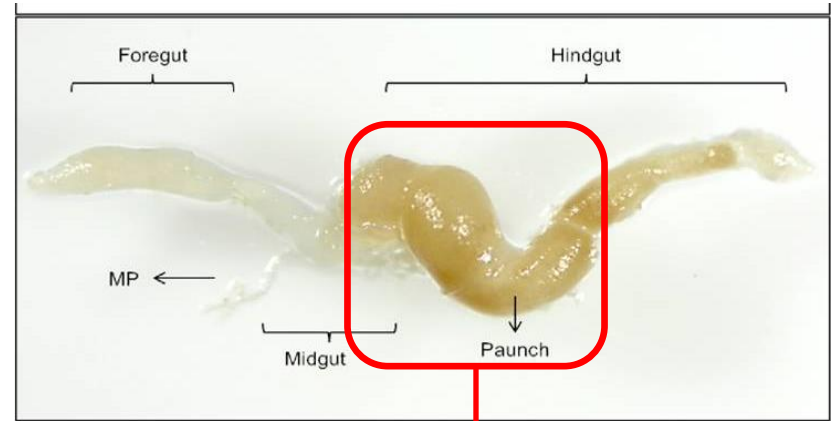
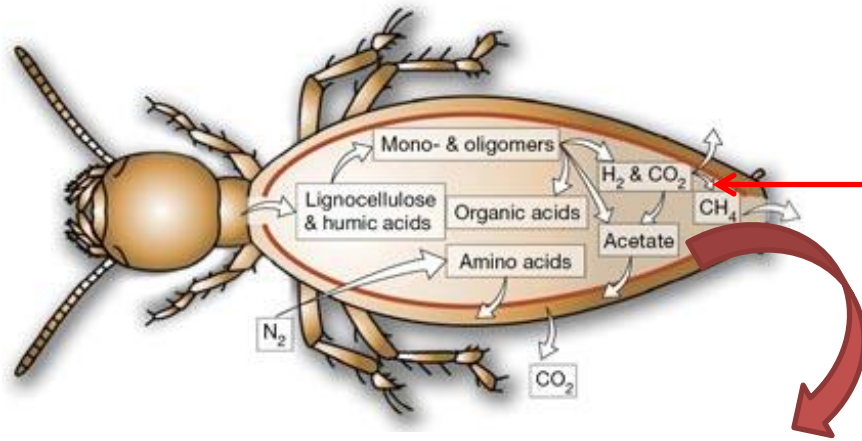
Termite

Adl et al. 2012

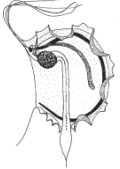
Bacteria



How do subterranean termites digest wood?



Who are the symbiotic flagellate protists?

					Flagellar apparatus/Mastigont system			Cytoskeletal arrangement			
	Closed mitosis	Golgi body	Hydrogenosomes	Nuclei	Flagellar system	Basal bodies arrangement	Microtubular fibers/sheets	Axostyle type	Undulating membrane	Costa type	Comb-like structure
 Hypotrichomonadea											
Parabasalia		+	+		Basically: 4 flagella	Basically: 4 basal bodies, 3 anterior; 1 recurrent	Pelta - covers the anterior part of the cell				
Class Hypotrichomonadea		+	+		4	biramous parabasal body		cone-like, usually stout	lamelliform	A	+
Class Trichomonadea		+	+		2 - 6	single karyomastigont		cone-like or reduced	lamelliform, if present	B, if present	-
Class Tritrichomonadea				1 or 2	0 - 5	single karyomastigont	pelta-axostyle complex may be reduced or absent	tube or cone-like, if present	rail-form, if present	A, if present	+/-
Class Cristamonadea				1 or more	2 to thousands per mastigont	single to multiple mastigonts	often spiralised or ramified axostyle	Stout rod or reduced	transformed into cresta (in some)	type A; reduced or absent	+ (ancestral) or absent
Class Trichonympha		+	+	1	hundreds to thousands	cell body divided into anterior rostrum and post-rostral area		thin and numerous, do not protude outside the cell	-	-	-
Class Spirotrichonympha		+	+	1		kinetosomes arranged into spiraled rows		tube-like and stout; multiple thin bands; or reduced	-	-	-
Preaxostyla											
Order Oxymonadida				1 or more	4, or a multiple of 4, flagella arranged in pairs	2 to several karyomastigonts	Preaxostylar lamina linking the 2 basal body pairs; Pelta (caps the nucleus)	crystalline axostyle; microtubular rod extending through the entire length of the cell	-	-	-

In Duarte et al. submitted

Hypotrichomonadea



Trichomonadea



Tritrichomonadea



Cristamonadea



Trichonympha



Spirotrichonympha



Oxymonadida

50µm

Objectives

H0 (null hypothesis)

“there are no differences among the symbiotic flagellate protists fauna living in the hindgut of termites which feed on different substrates”

- comparison of flagellate protists communities in terms of diversity and abundances
- high quality microscopy images of flagellate protists
- 1st assessment of possible methods to perform a molecular analysis of flagellate protists



From: <http://sostermitas.angra.uac.pt/termitas/>

Methods

Termite collection

Laboratory trials

Termite dissection

Protist counting

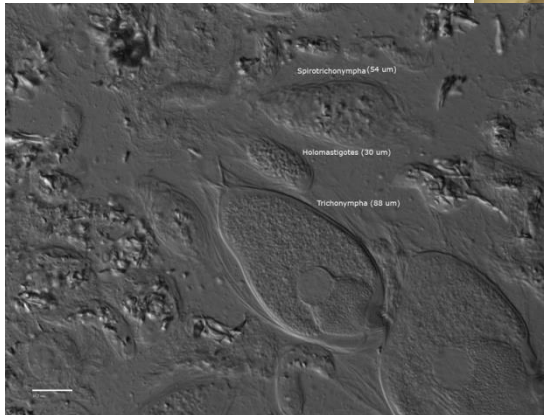
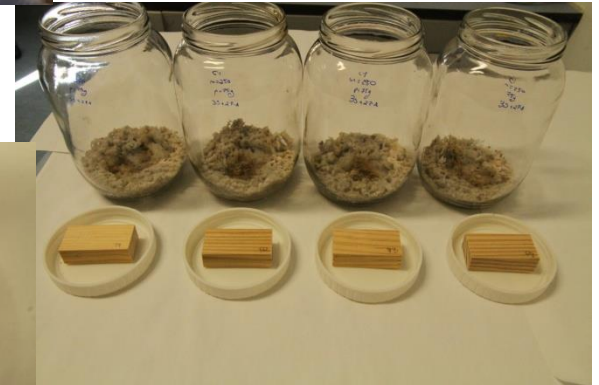
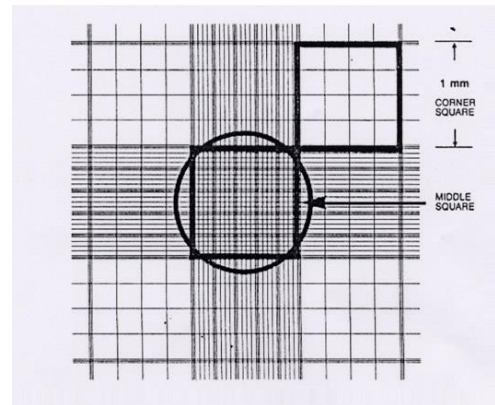


Image (DIC) by Dan Gage



Haemocytometer

...before STSM.

Methods

Diets

- **Natural diet** – pieces of the wood from where termites were collected
- **Pine** – *Pinus pinaster* Ait.
- **Beech** – *Fagus sylvatica* L.
- **Beech TMT** – beech submitted to a thermal treatment of 180°C for 4h
- **Cellulose** – powder cellulose plus deionized water
- **Starving** – no cellulose source offered to the termites

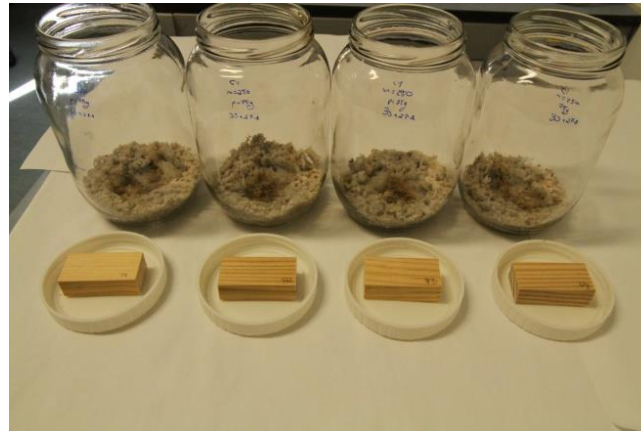


...before STSM.

Methods

EN117 (adapted)

14 days of exposure
(except for starving =
6 days)



3 replicates for each diet
(except for natural and
starvation – 1 replicate)

250 termite workers (*Reticulitermes
grassei* Clément) per trial

...before STSM.

Methods

Preparation methods for microscopy analysis

Step	Solution	Duration	Date
1 Fixation	1.25% glutaraldehyde, 2% paraformaldehyde in 0.05% Pipes buffer	fixed in Lisbon (4); overnight (1; 3)	19-08-2014
2 Wash	Passende buffer - 0.1M CaCodylat buffer, pH7.4	10 min (x3)	20-08-2014
3 Post Fixation	1% OsO4 in 0.1M CaCo	90 min	20-08-2014
Wash	0.1M CaCo, pH7.4	10 min (x3)	20-08-2014
4 Dehydration	70% EtOH	10 min	20-08-2014
	90% EtOH	10 min	20-08-2014
	96% EtOH	10 min	20-08-2014
	100% EtOH	10 min (x4)	20-08-2014
5 Infiltration	1 LR White : 3 100% EtOH	overnight	20-08-2014
	1 LR White : 1 100% EtOH	3h	21-08-2014
	3 LR White : 1 100% EtOH	3h	21-08-2014
	100% LR White	overnight	21-08-2014
6 Embedding	100% LR White, at 60°C	overnight	22-08-2014

Preparation and embedding in resin of samples for TEM analysis

For each variable – 30 termites hindguts prepared

DNA and RNA tentative extraction - X



@Norwegian Forest and
Landscape Institute

...during STSM.

Methods

Ultramicrotome



Leica EM UC6

Transmission Electron Microscope



FEI Morgagni 268



@Imaging Centre of the Norwegian
University of Life Sciences, NMBU

...during STSM.

Results

Morphotypes ID

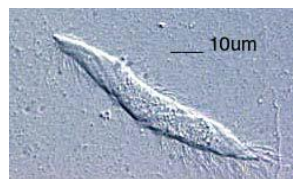
	Phylum	Class	Order	Family	Genus	Species
n1	Parabasalia	Trychonymphea	Trichonymphida	Trichonymphidae	<i>Trichonympha</i>	<i>Trichonympha agilis</i>
n2	Preaxostyla	-	Oxymonadida	-	<i>Pyrsonympha</i>	
n3	Preaxostyla	-	Oxymonadida	-	<i>Dinenympha</i>	<i>Dinenympha gracilis</i>
n4	Parabasalia	Spirotrichonymphea	Spirotrichonymphida	Holomastigotoididae	<i>Holomastigotes</i>	<i>Holomastigotes elongatum</i>
n5	Preaxostyla	-	Oxymonadida	-	<i>Dinenympha</i>	<i>Dinenympha fimbriata</i>
n6	Parabasalia	Hypotrichomonadea	Hypotrichomonadida	Hypotrichomonadidae	<i>Trichomitus</i> or <i>Trichomitopsis</i>	
n7	Preaxostyla	-	Oxymonadida	-	<i>Pyrsonympha</i>	
n8	Parabasalia	Spirotrichonymphea	Spirotrichonymphida	Holomastigotoididae	<i>Microjoenia</i>	
n9	Parabasalia	Trychonymphea	Trichonymphida	Teranymphidae	<i>Pseudotrichonympha</i>	
n12	Parabasalia	Spirotrichonymphea	Spirotrichonymphida	Holomastigotoididae		
n13	Parabasalia	Trichomonadea	Honigbergiellida	Tricercomitidae	<i>Tricercomitus</i>	
n16	Preaxostyla	-	Oxymonadida	-		
n18	Parabasalia					



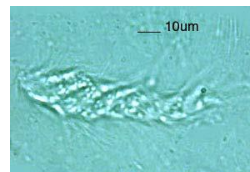
T. agilis



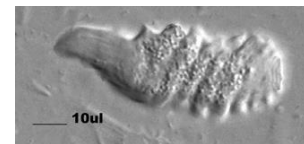
Microjoenia



D. fimbriata



H. elongatum

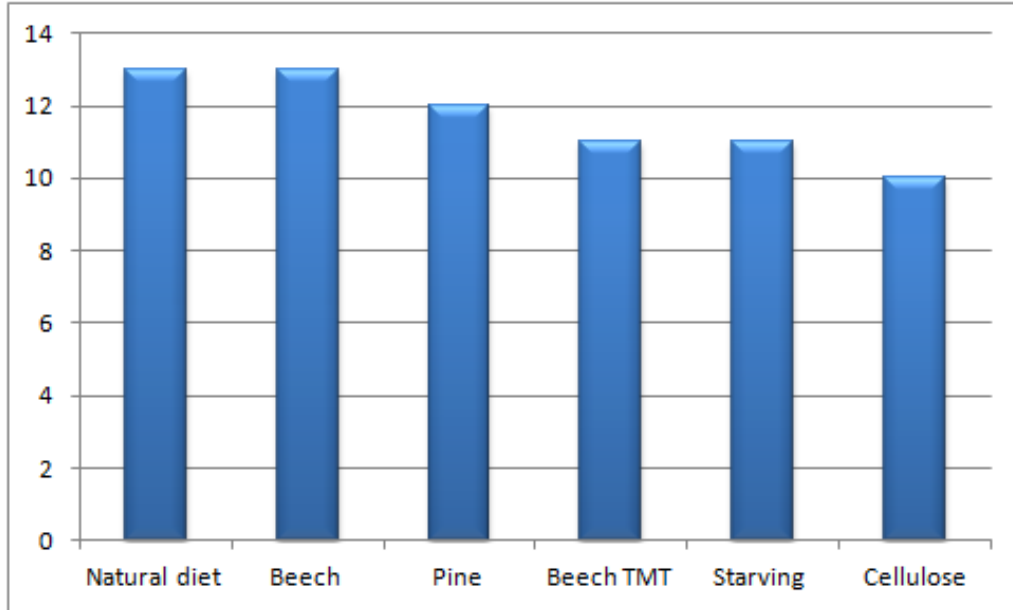


Pyrsonympha spp.

Photos of JL Lewis

Results

Number of morphotypes



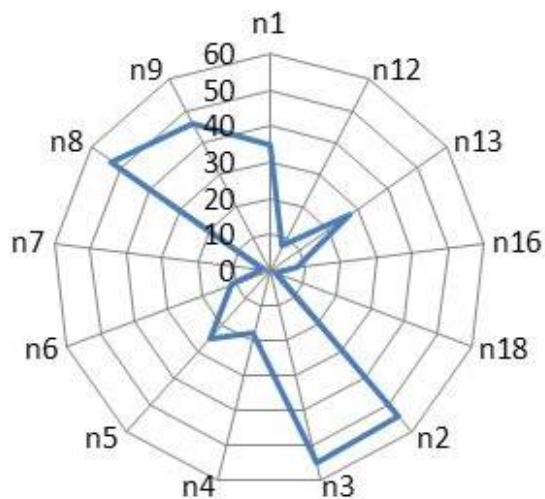
	Survival rate (%)	Mass loss (%)
Natural diet	74.4	5.2
Pine	39.3	3.7
Beech	45.9	3.6
Beech TMT	39.5	3.7
Cellulose	39.6	11.4
Starving	33.2	-

Results of paired T-test ($p < 0.05$)

	Natural diet	Pine	Beech	Beech TMT	Cellulose
H0? Natural diet					
Pine	✓				
Beech	✓	✓			
Beech TMT	✗	✗	✗		
Cellulose	✗	✗	✗	✓	
Starving	✗	✗	✗	✓	✓

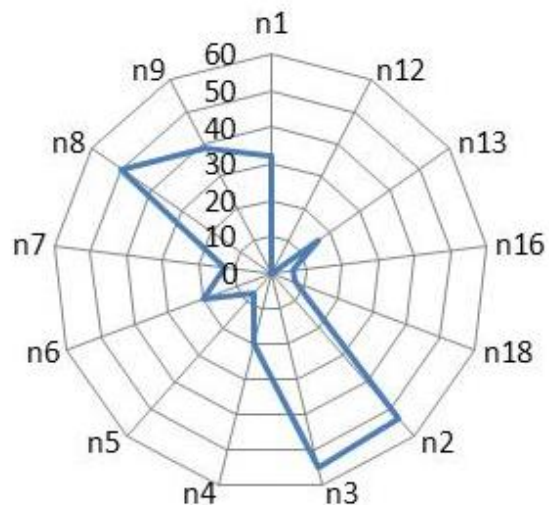
Natural diet

↑ *Dinenympha fimbriata* – n5

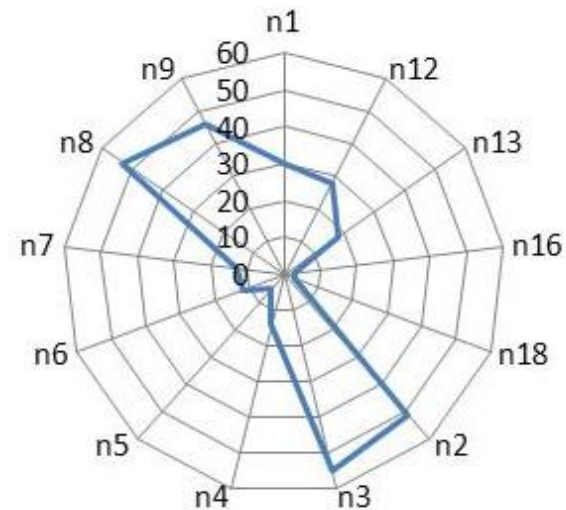


Pine

↑ *Hypotrichomonadidae* – n6

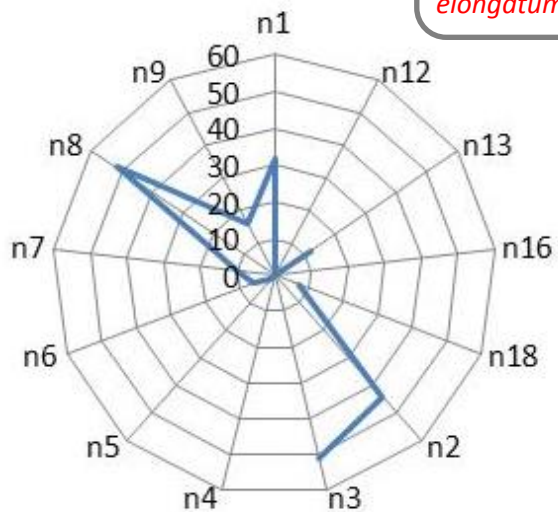


Beech



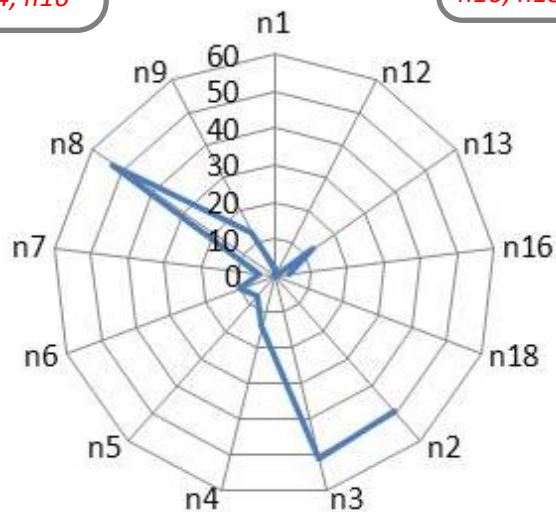
Cellulose

↓ *Pseudotriconympha* spp. – n9
Holomastigotes elongatum – n4; n16



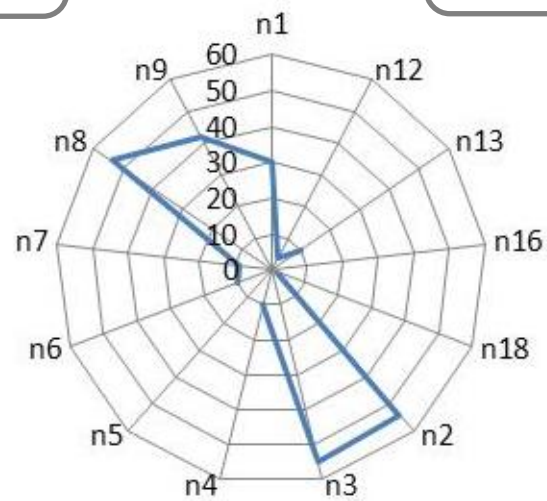
Starving

↓ *Trichonympha agilis* – n1
n16; n18



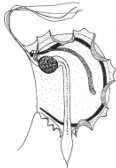
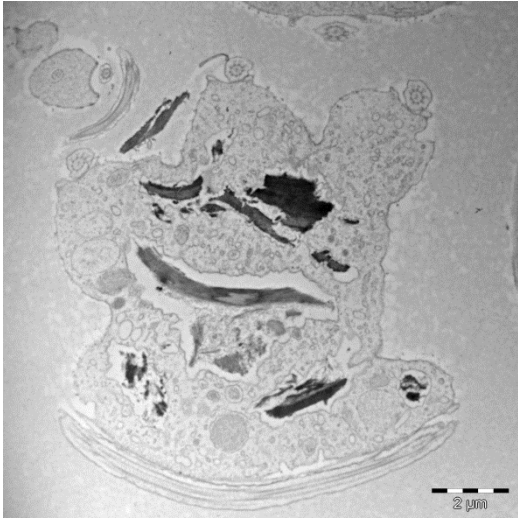
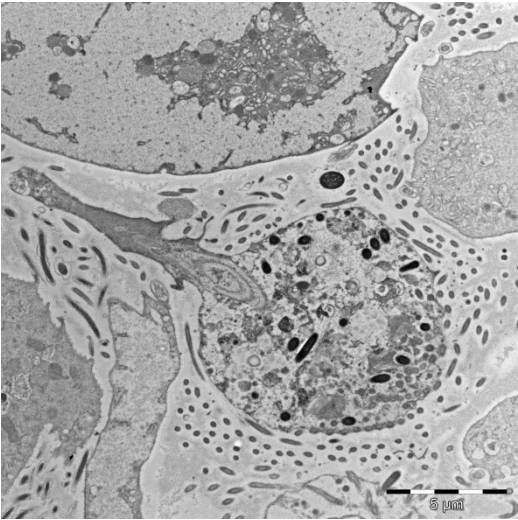
Beech TMT

Dinenympha fimbriata – n5
n16; n18



Results

Parabasalia



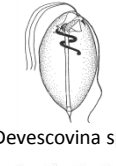
Trichomitopsis spp.



Trichomitus spp.



Triitrichomonadidea



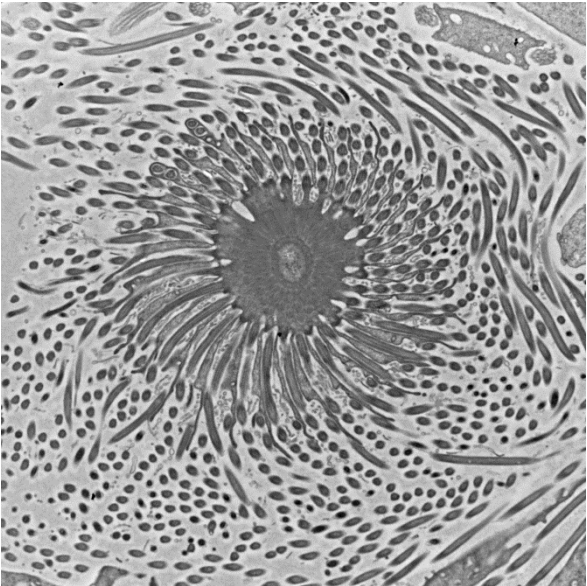
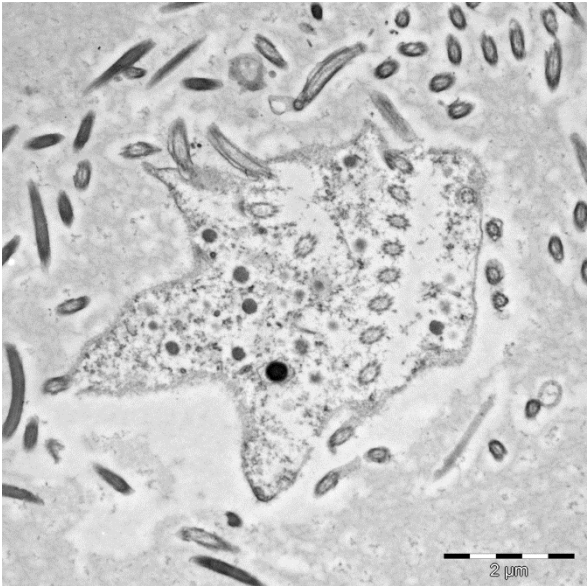
Devescovina spp.



Trichonympha spp.

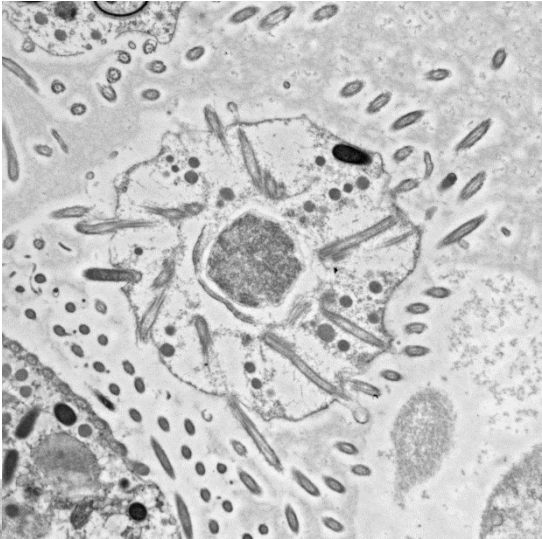


Holomastigotes spp.

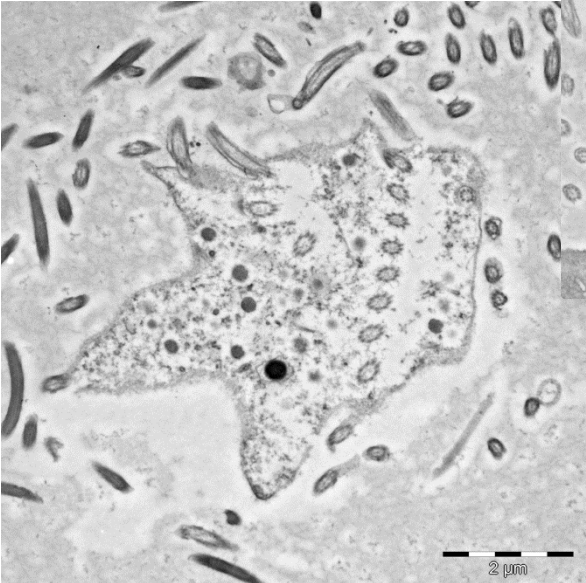
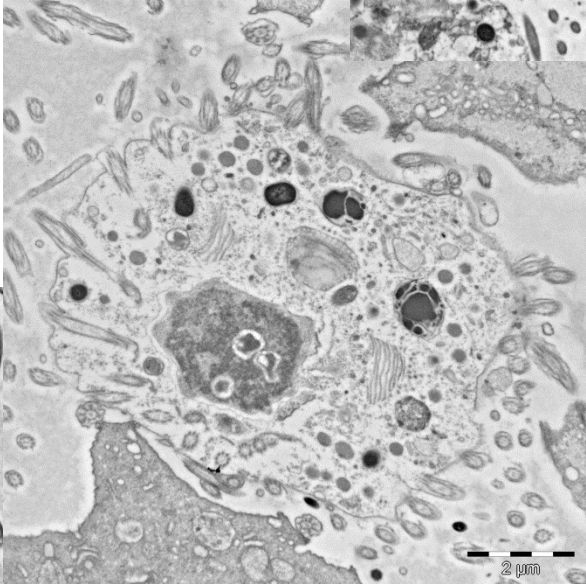


Results

Parabasalia



Spirotrichonymphidae

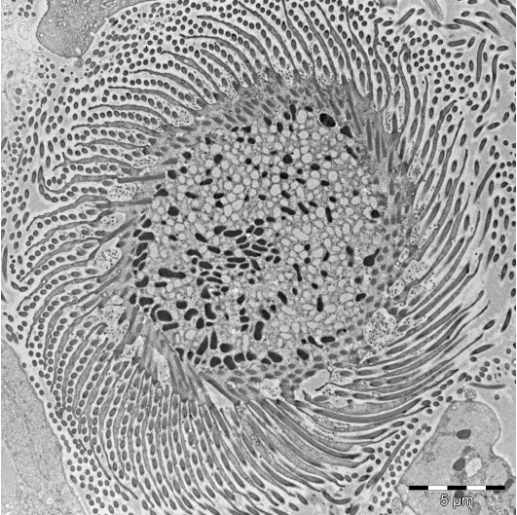
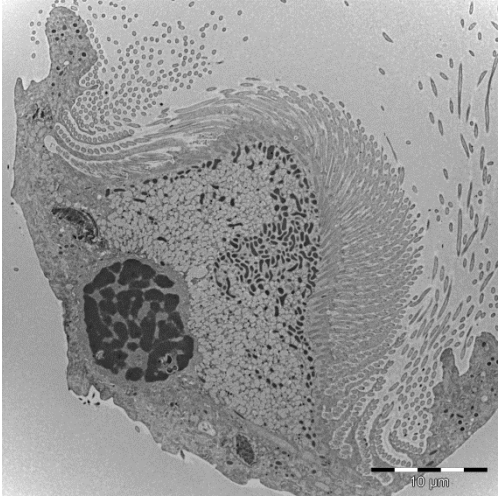
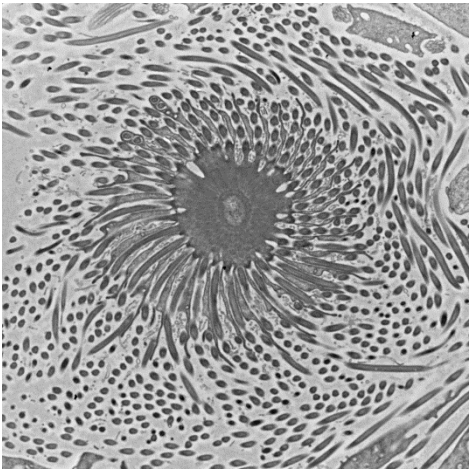
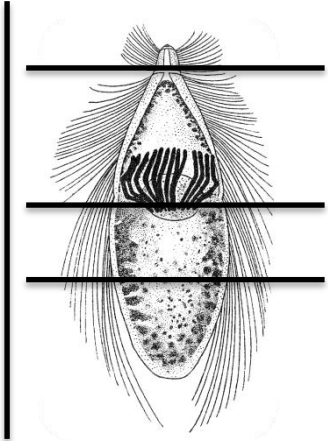
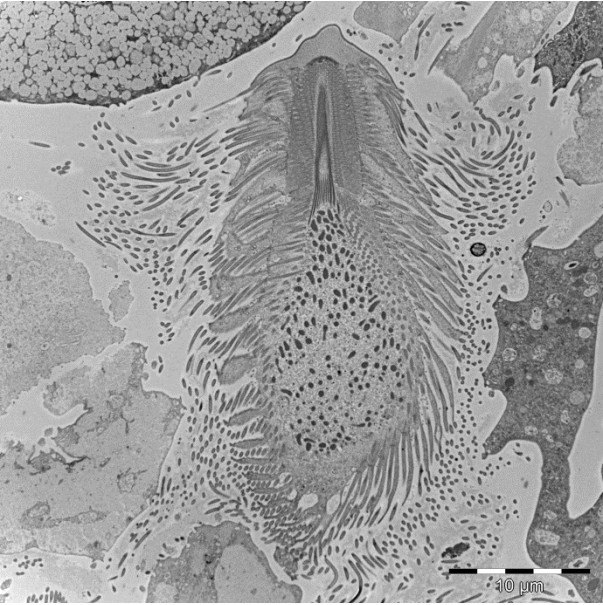


Flagella - spiralized pattern



Results

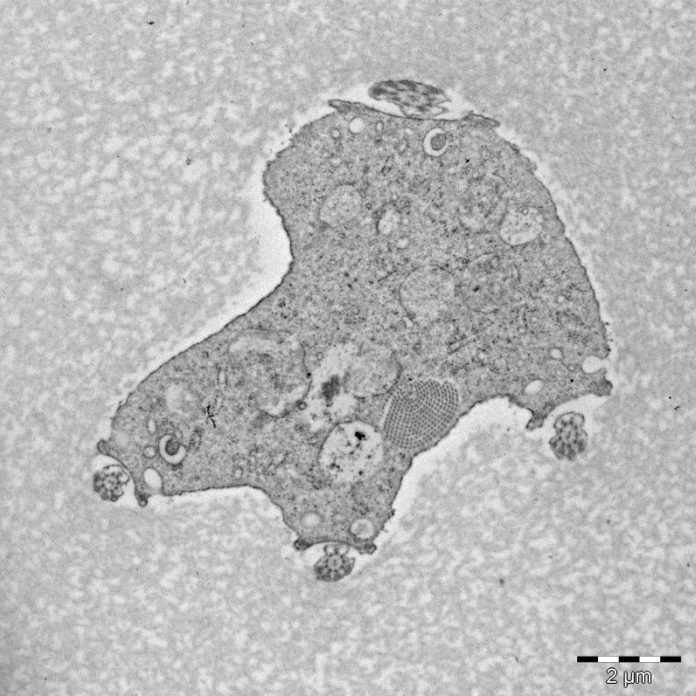
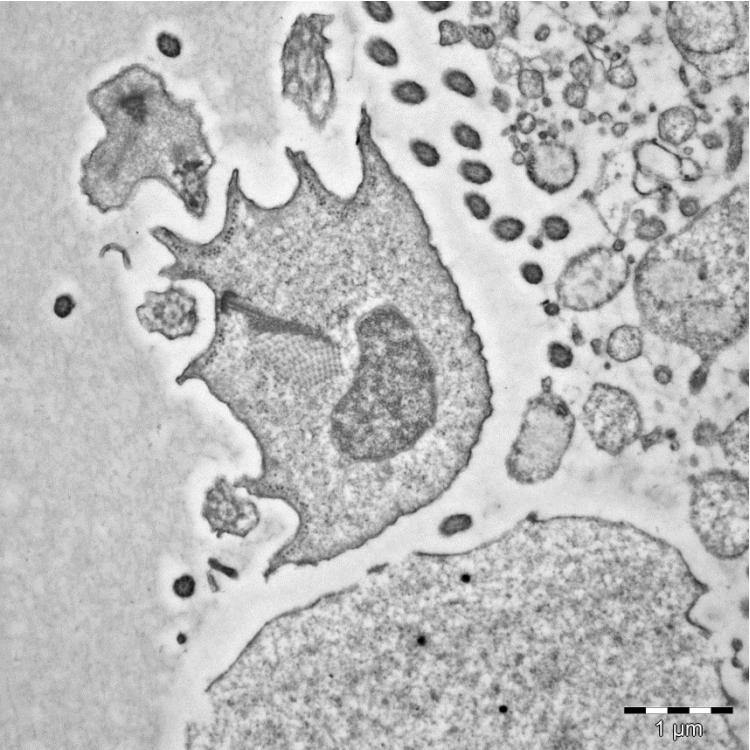
Parabasalia



Trichonymphidae

Results

Preaxostyla



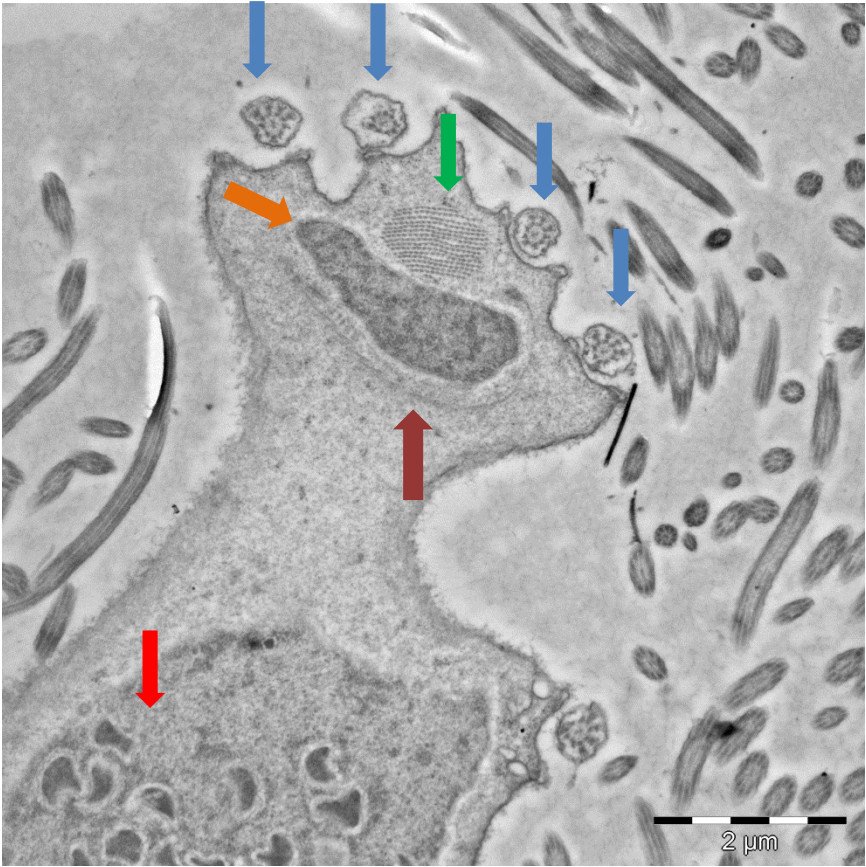
Pyrsonympha spp.



Oxymonas spp.

Results

Preaxostyla



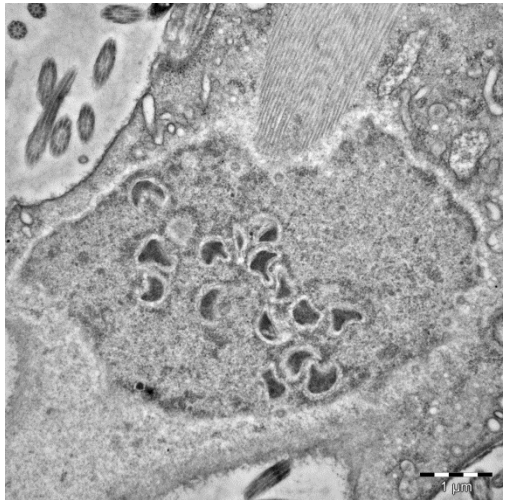
4 Flagella

2nd Nucleus

Axostyle

Pelta

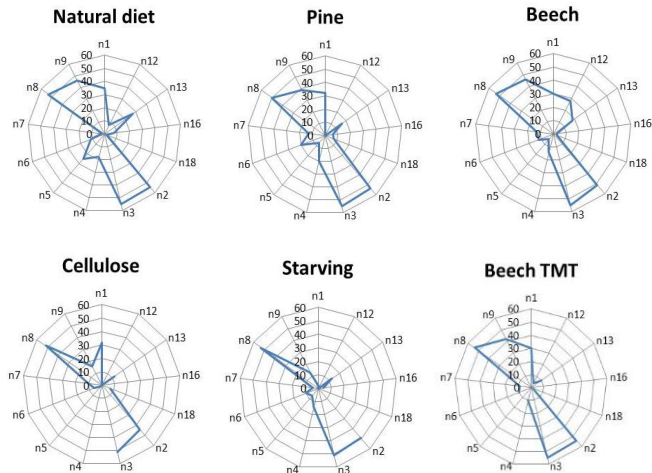
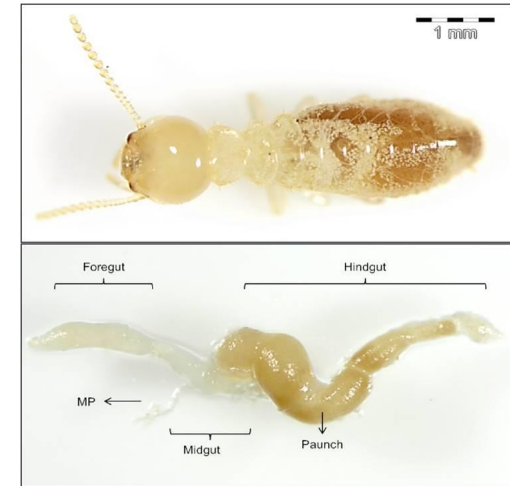
Nucleus w/ intranuclear symbionts



Discussion/Conclusions

There are significant differences in the flagellate protists community of termites according with different type of diets

This work was a first step towards the understanding of the changes occurring inside *Reticulitermes grassei* hindgut due to the ingestion of different diets



The evaluation of the flagellate protist community may be a valuable tool to detect substances which may constrain the termite hindgut “bioreactor” correct functioning

Acknowledgments

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Hilde Kolstad

Trine Hvoslef-Eide



Nina



Elin, Trine and Hilde



Inger

**Thank you for your
attention**

